

Design of an innovative photobioreactor stirred by fractal oscillating grids for microalgae culture – Determination of the mixing time

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Green microalgae Red microalgae Cyanobacteria Haptophyta Dinoflagelate Diatoms

Shear stress sensitivity

Microalgae culture aims: CO₂ capture and absorption, production of interest products as lipids (which can be transformed into biodiesel), photosynthetic pigments, proteins, vitamins or carbohydrates used for therapeutic, food or cosmetic purposes^{1,3,4,5}.

| | Simple cartesian | Fractal cartesian | Fractal square pattern | I-fractal pattern | |
|----------------------------|------------------|-------------------|---------------------------|---------------------|---------------------------|
| | pattern | pattern | | Square fitting grid | Rectangle fitting grid |
| Pattern design | | | | | |
| olidity | 0.3 | 0.3 | 0.3 | 0.3 | 0.3 |
| Ν | 2 | 2 | 4 | 3 | 3 |
| ₀ [mm] | 234 | 234 | 132.9 | 140.8 | 145.1 |
|) [mm] | 5.5 | 9.5 | 8.7 | 4.9 | 12.1 |
| ratio _L | 0.49 | 0.48 | 0.5 | 0.45 | 0.43 |
| ratio _l | 1 | 0.5 | 0.5 | 0.9 | 0.53 |

Principle of the PLIF method

The first experiment for the PBR characterisation is the **determination of the mixing time** according to the grid geometry and the oscillation parameters (ν , A, z_{eq}). This kind of experiments are performed by a Planar Laser Induced Fluorescence (PLIF) method: the dispersion of a fluorescent dye (Rhodamine 6G) in the fluid medium (water) illuminated by a laser sheet (generated by a continuous laser Dantec Dynamics RayPower) is record by a scientific camera (LaVision Imager M-lite 5M) with an optical filter cutting the laser radiation off.



The local fluorescence radiation is linked to the local dye concentration, the laser radiation power and the optic set-up.

 $I_{\text{fluo}} = \eta_{\text{optic}} \cdot I_{\text{laser}} \cdot e^{\varepsilon \cdot L \cdot C} \cdot \eta_{\text{quantum}} \cdot V \cdot C$ \Rightarrow Calibration curves $I_{\text{fluo}}(C)$

C depends on the location (x and z) on a considered picture \Rightarrow calibration pixel by pixel. Due to the vertical movement of the grid, the calibration curves depend on the grid position z_{grid} too \Rightarrow grid tracking algorithm.





Determination of the mixing time according to the oscillation parameters

The calibration of the PLIF is limited in our case due to saturation and Beer-Lambert absorption resulting in a hook effect. Instead of dealing with calibration, we propose the use of the normalised pixel intensity *I*norm to overcome the heterogeneity of the pixel intensity due to the laser illumination.

$$I_{\text{norm}} = \frac{\left| I\left(x; z; z_{\text{grid}}\right) - I_0\left(x; z; z_{\text{grid}}\right) \right|}{\left| I_{\infty}\left(x; z; z_{\text{grid}}\right) - I_0\left(x; z; z_{\text{grid}}\right) \right|} \qquad \begin{array}{l} \text{Before injection,} \\ I_{\text{norm}} \to 0 \end{array}$$

$$\begin{array}{l} \text{After the injection,} \\ I_{\text{norm}} \to 1 \end{array}$$

With I_0 the considered pixel intensity before the dye injection and I_{∞} the considered pixel intensity when the dye concentration homogeneity is reached.



Project perspectives

The evaluation of the mixing time by the PLIF method seems to be subject to a great variability. This variability. This variability. This variability can be attributed to the presence of a non-repeatable mean flow, the presence of which may or may not be confirmed by PIV (Particle Imaging Velocimetry) measurements (scheduled during the last quarter of 2022 to complete the hydrodynamic study). The results of the mixing time determination study will give us a tendency of which grid is the best for a mixing application, even if this will not be, in our opinion, the most discriminating performance criterion for microalgae culture. To identify the areas of high shear stress (which we believe is the most critical parameter) in the PBR, experiments using the bioluminescence ability of the dinoflagellate (shear-sensitive microalgae) Pyrocystis fusiformis will be developed and performed. All the results of the hydrodynamic and turbulence study will be used to select the, a priori, best configurations to perform microalgae cultures and evaluate the culture performances of the fractal oscillating grid stirred PBR. Besides all this, DNS (Direct Navier-Stokes) simulations will be performed with OpenFoam to simulate the flows generated by fractal oscillating grid systems. The simulation results will be compared to the experimental PIV results.

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